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# Recent benthic foraminiferal assemblages in deep high-energy environments from the Gulf of Cadiz (Spain)

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## Abstract

The benthic environment in the Gulf of Cadiz, north-eastern Atlantic, is strongly affected by the Mediterranean outflow water undercurrent (MOW) which flows northwards along the western Iberian Margin at 500–1500 m water depth. Foraminiferal census counts of living and dead assemblages from 27 surface samples ranging from 103 to 1917 m water depth, and the examination of hard substrates reveal a close correlation of the fauna with the local hydrography and sediment facies. Four different faunal groups are separated by factor analysis of the living fauna. Assemblage 1 contains typical lower slope species and dominates samples from the lower MOW core layer and in the North Atlantic deep water below. Shelf edge foraminifera are common in assemblage 2a which shows the highest proportions in samples from 103 to 272 m. Assemblage 2b is dominated by upper slope species and suspension-feeders that are frequent in the upper MOW core layer and in distal settings between 396 and 901 m. Species from assemblage 3 prefer epibenthic habitats and are recorded with high proportions exclusively in the immediate flow paths of the upper MOW between 496 and 881 m. Colonisation structures and species composition of epibenthic assemblages from the proximal facies largely differ from those in distal settings. In general, epibenthic foraminifera only use elevated substrates under the influence of near-bottom flow. Under high current velocities, epibenthic foraminifera prefer large and heavy objects. They colonise high attachment levels where a maximum yield of advected food particles can be achieved. In distal settings at lower flow velocities, the elevation height does not exceed 20 mm above the surrounding sediment surface. This level is related to a hydrologic transition layer with high concentrations of suspended particles. The comparison of microhabitat preferences and faunal structure under high and low current velocities reveal that substrate stability may be a confining environmental variable for endobenthic and shallow epibenthic foraminifera. The observations also indicate that the preferential settling height of epibenthic foraminifera is related to the highest lateral flux rates of food particles within reach from the sea floor. A dynamic selection of elevated microhabitats is only used by 7.8% of all species recognised in the Gulf of Cadiz area. © 2002 Elsevier Science B.V. All rights reserved.

*Keywords:* benthic foraminifera; epibiotism; continental slope; East Atlantic Ocean

## 1. Introduction

Benthic foraminifera are widely used for pa-

laeoenvironmental reconstruction. Before application to the fossil record, index species, assemblages or faunal indices have to be calibrated with scaling ecological parameters. A growing number of studies on the recent distribution of living (Rose Bengal-stained) benthic foraminifera prove

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a substantial relationship with the flux of particulate organic matter to the sea floor (Lutze and Coulbourn, 1984; Altenbach and Sarnthein, 1989; Herguera and Berger, 1991; Gooday, 1994; Altenbach et al., 1999; Loubere and Fari-duddin, 1999), the oxygen content of ambient bottom waters (Koutsoukos et al., 1990; Hermelin, 1992; Alve, 1995; Bernhard and Sen Gupta, 1999), and high levels of environmental stress (Sharifi et al., 1991; Yanko et al., 1994; Alve, 1995; Hess and Kuhnt, 1996; Stott et al., 1996).

Water turbulence leaves also a certain impact on benthic foraminiferal assemblages. This has been demonstrated indirectly in most cases by a coincidence of changes in assemblage composition at the depth of the wave base and the shelf break where water turbulence markedly decreases (Pujos, 1971; Pujos-Lamy, 1973; Seiler, 1975; Lutze, 1980; Schiebel, 1992; Levy et al., 1993; Debenay and Redois, 1997). In shallow-water, high-energy environments, lateral displacement of foraminiferal tests is common and obstructs environmental interpretations if the living, autochthonous fauna is not considered (Murray, 1970; Murray et al., 1982; Williamson et al., 1984).

Little is known about foraminiferal assemblages in deep high-energy environments. As in shallow waters (Dobson and Haynes, 1973; Haward and Haynes, 1967; Semeniuk, 2000), epibenthic habitats are used with greater abundances under the impact of near-bottom flow regimes (Mullineaux, 1988; Schönfeld, 1997, 1998), and suspension-feeders are frequent in adjacent places where water turbulence decreases and suspension load settles (Lutze and Altenbach, 1988; Mackensen et al., 1985; Thomsen, 1998). The patchy and sparse distribution of appropriate elevated habitats (Oschmann, 1990) may lead, if not sampled or analysed, to a misinterpretation of the dead assemblage in the near-surface sediment (Mackensen, 1987). Nonetheless, the observations made to date rely only on a few tens of samples from selected areas. The deep, high energy facies deserves attention, however, because it is well established in sea straits which provide pathways for inter-basinal faunal exchange and deep-water circulation.

In the present paper, I demonstrate how the

benthic foraminiferal assemblage composition is structured by the influence of the Mediterranean outflow water (MOW) contour current in the Gulf of Cadiz, Spain. Observations of microhabitat colonisation structures reveal the influence of substrate properties, current strength, and the strategies of foraminifera to prevail in this deep high-energy environment.

## 2. Study area and hydrography

The Gulf of Cadiz forms a southwest-facing, arched embayment of the north-eastern Atlantic ocean between the Gibraltar Strait and Cape San Vicente. The shelf is covered with sand and rock outcrops in places with strong exposure, in particular south of the Algarve and to the east of Cadiz up to Gibraltar (López-Galindo et al., 1999). Extended mud drapes occur on the shelf close to the river mouths of Huelva and Guadalquivir, and in the inner part of the Gulf of Cadiz (Nelson et al., 1999). Glauconitic sands prevail at the shelf edge and pass into muds with intercalated sandy contourite beds on the upper slope (Heezen and Johnson, 1969; Faugères et al., 1984; Sierro et al., 1999). The shelf environment is under influence of North Atlantic Central Water (NACW) from the thermocline (approximately 30–100 m) down to 430 and 600 m depth at maximum (Wüst, 1936). The flow regime in the near-surface water is from northwest to southeast (Lobo et al., 2000).

The MOW is present between 450 and 1500 m. It is the most outstanding hydrographic component in this area and shows higher temperatures and salinities than the NACW (Heezen and Johnson, 1969; Zenk, 1975a). The MOW enters the Gulf of Cadiz through the Gibraltar Strait at depths of 150–280 m (Lacombe et al., 1964). On entering the Atlantic, it cascades down to 700 m depth and rapidly mixes with ambient NACW (Zenk, 1971, 1975b; Ambar and Howe, 1979a). Then it turns clockwise closely following the slope in a narrow vein that divides and reunites again further downstream (Madelain, 1970). The MOW's buoyancy is in disequilibrium with the ambient NACW and it is held upwards only by

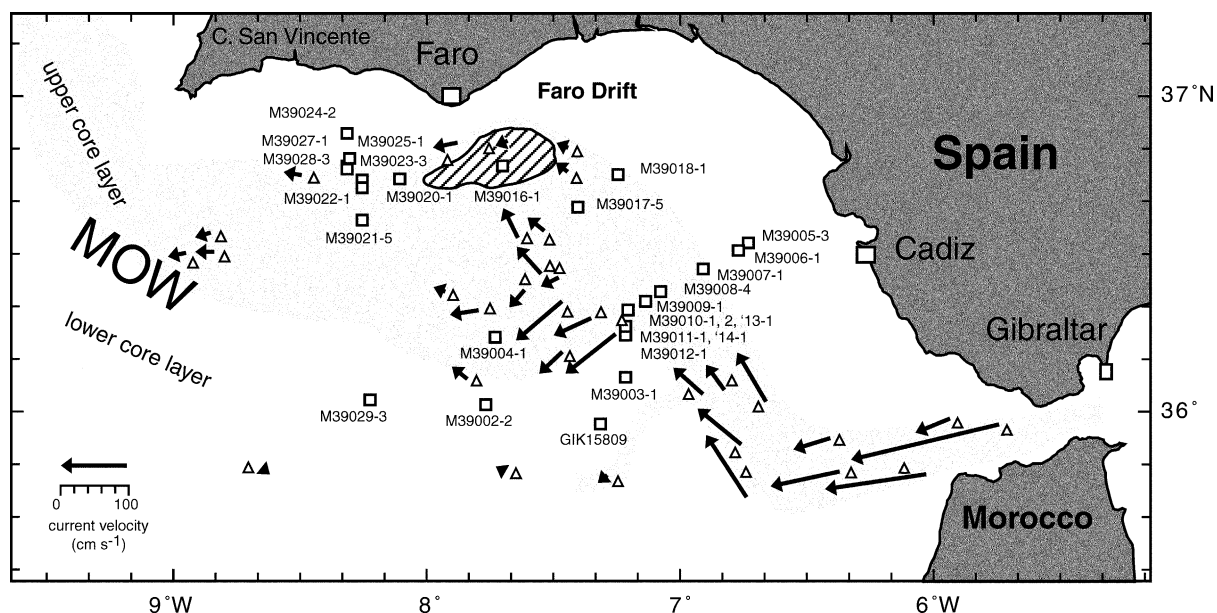


Fig. 1. Location of sampling sites (open squares) in the Gulf of Cadiz, north-eastern Atlantic. The spreading of the MOW is depicted in light and medium grey. The MOW flow strength and direction as obtained from current meter moorings (open triangles) are marked with arrows (after Heezen and Johnson, 1969; Madelain, 1970; Zenk, 1975b; Zenk and Armi, 1990; Nelson et al., 1993). The Faro Drift (hatched area) is given for comparison (Faugères et al., 1984).

Coriolis force (Baringer and Price, 1997). When the current is slowed down at slope-perpendicular ridges or at canyons, the outflow water further descends and spreads at its equilibrium density level between 1000 and 1400 m water depth forming the lower MOW core layer (Zenk, 1975b; Thorpe, 1976; Gründlingh, 1981; Baringer and Price, 1999). Close to the Gibraltar Strait, the current speed is rather high, up to  $200 \text{ cm s}^{-1}$  (Bøyum, 1963; Lacombe et al., 1964; Kinder and Bryden, 1987). In the northern branch, the current in the upper core layer slows down to  $100 \text{ cm s}^{-1}$ , and the velocity further decreases to about  $13 \text{ cm s}^{-1}$  in distal settings off Cape San Vicente (Meincke et al., 1975; Ambar and Howe, 1979b; Nelson et al., 1993). The currents are slow in the lower core layer, where only  $5\text{--}10 \text{ cm s}^{-1}$  are recorded (Heezen and Johnson, 1969).

The bottom sediments in the MOW depth interval also reflect the decrease in current strength from east to west. Hard rock outcrops and gravel lags occur in proximal areas in the east (Heezen

and Johnson, 1969; Nelson et al., 1993). Sand and sandy silt covers the sea floor along the main flow paths, and silt and clayey silt prevails in quiet zones outside and in between the MOW veins (Nelson et al., 1999). The MOW flow has also resulted in the build up of the Faro Drift and other sediment drift bodies in the west that consist of sandy to silty sediment accumulated and sorted under the current regime (Fig. 1) (Gonthier et al., 1984; Faugères et al., 1984, 1999; Stow et al., 1986).

Below the MOW, North Atlantic deep water is present and shows only slow movements (Zenk, 1980). The bottom sediments pass from hemipelagic silty clays into foraminiferal oozes (Lebreiro et al., 1997).

Summer upwelling due to persistent northerly winds is a common feature of the entire western Iberian coast (Abrantes, 1988). Intensity maxima are recognised in the coastal regions off Galicia ( $42\text{--}44^\circ\text{N}$ ) and off Cape Sines at  $38^\circ\text{N}$  (Fiúza, 1983; Fiúza et al., 1982). In the Gulf of Cadiz, seasonal upwelling only takes place in the north-

ern part, off the Algarve coast between Cape San Vicente and Faro. Satellite sea surface temperature images processed by the Plymouth Marine Laboratory Remote Sensing Group indicate that upwelling off the Algarve seldomly reaches intensities as off northern Iberia (e.g. [http://www.npm.ac.uk/rsdas/composite/po\\_comps/full\\_size/weekly\\_all/1998/](http://www.npm.ac.uk/rsdas/composite/po_comps/full_size/weekly_all/1998/)). Satellite data reveal that ocean surface productivity ranges from 153 to 219 g C m<sup>-2</sup> yr<sup>-1</sup> in the Gulf of Cadiz (Sathyendranath et al., 1995) while earlier compilations of primary productivity maps allow an estimate of 82 g C m<sup>-2</sup> yr<sup>-1</sup> for the entire Cadiz region (Berger et al., 1987). Later approaches reckon 90–150 g C m<sup>-2</sup> yr<sup>-1</sup> from Coastal Zone Colour Scan remote sensing data (Antoine et al., 1996).

### 3. Material and methods

#### 3.1. Foraminiferal studies

Twenty-seven sediment surface samples from the Gulf of Cadiz were used in this study. They were recovered with a large box corer with 50 × 50 cm surface area and a Van Veen grab sampler of 45 × 45 cm size during RV *Meteor* cruise M39 in April and May 1997 (Schott et al., 1999). One sample (GIK15809) from RV *Noroit* cruise KR 84 in 1984 was also analysed. That sample was also recovered with a large box corer.

Macrobenthos and other hard substrates like mollusc shells, ships clinker, deep-water corals, hydroids, and sponges were carefully removed from the sediment surface of box cores and conserved in methanol. These objects were later examined for attached epibenthic foraminifers. The maximum height of the substrates or points of attachment above the ambient surface was recorded. For massive objects as pebbles and shells that were partly embedded in the surface sediment, zero level for attachment height was assigned to a horizon from where below traces of fine sediment were still sticking to the object. For free-standing, delicate substrates as hydroids, *Rhabdammina* tubes or *Pheronema* sponges, a 'mudline' of fine, adherent sediment was commonly recognisable.

A volume-defined sample from the uppermost centimetre of the surface sediment was taken and immediately conserved and stained with a solution of 2 g Rose Bengal in 1 l methanol p.a. (Lutze and Altenbach, 1991). This sample was used to study the living, shallow epi- and endobenthic foraminifers, and the dead assemblage in the surface sediment. The surface area sampled was 173–260 cm<sup>2</sup> on box core surfaces and 87–150 cm<sup>2</sup> on grab sample surfaces. A representative coverage of different microenvironments on the sediment surface was attempted wherever possible.

The samples for foraminiferal studies were washed on a 63-µm mesh screen. They were first passed through a 2000-µm sieve to collect larger particles and fragile tubular arenaceous foraminifers which can be damaged if washed too long. The dried residues were further divided into the grain-size fractions 63–250 µm, and 250–2000 µm. Benthic foraminifers were analysed from the fraction > 250 µm to exclude smaller forms which are often displaced by redeposition (Lohmann, 1978; Lutze, 1980), and to make the data comparable to previous benthic foraminiferal studies in adjacent areas (Lutze and Coulbourn, 1984; Schönfeld, 1997). The samples were then picked for well-stained benthic foraminifers that are considered to have been living at the time of sampling (Murray and Bowser, 2000). From sand-rich samples (e.g. M39009-1 and M39023-3), splits of up to 1/8 were made to facilitate microscopic work. Arenaceous, and thick-walled miliolid species were either soaked with water to make their staining pattern more visible or crushed to see whether the test contained protoplasm. After the living forms were sorted out, a further split was made and picked for dead foraminifers. Living and dead assemblages were collected in Plummer cell-slides, sorted at species level, fixed with glue, and counted.

Multivariate statistical analyses were applied to the faunal census data by using StatView 4.02© Abacus Concepts, Berkely, CA, USA, for Macintosh®. Species were grouped by factor analysis with principal component solution and varimax rotation.

## 4. Results and discussion

### 4.1. Foraminiferal assemblages in surface sediments

The foraminiferal fauna shows middle to upper bathyal characteristics and is very similar to assemblages previously described from other adjacent parts of the eastern North Atlantic (Calvez, 1958; Colom, 1952; Seiler, 1975; Lutze, 1980; Levy et al., 1995; Schönfeld, 1997). 192 different species are recognised, of which 74 are Textulariina, 30 are Miliolina and 88 are Rotaliina. To reduce the high species number to a practical size, only those species which occur in at least one sample among the first five ranked species were taken into consideration. This results in a reduction to 58 and 57 species which comprise 67.3–87.0% and 64.9–89.7% of the living and dead assemblage, respectively. Taxonomic references and the percentages of living and dead benthic foraminiferal taxa that are used for the multivariate analyses are given in Appendices 1–3.

Factor analyses were applied to the reduced data sets in order to obtain species associations with distinct depth distributions or demands to specific environmental conditions. Living and dead assemblages were not combined but analysed separately. Foraminiferal species were assigned to different associations following the analysis of the living assemblage. The living fauna is considered to be in equilibrium with the ambient environmental parameters and not biased by re-deposition, postmortal loss of arenaceous species, and degradation due to bioerosion (Murray et al., 1982; Schröder, 1986; Martin and Liddell, 1991). For 58 frequent benthic foraminiferal species from 21 samples where the living assemblage has been assessed, the analysis extracts 65 factors. The four first ranked factors are clearly separated from the others by their variance proportion, and they account for 46.7% of the total variability of the data set (Table 1). The best resolution of faunal groups is achieved between factors 1 and 2. Factor loading of species infer three main clusters that are assigned to Assemblages 1, 2 and 3 (Fig. 2). Assemblage 2 may be subdivided further into assemblages 2a and 2b.

Species of Assemblage 1 show high, positive factor 1 loading ( $>0.243$ ) and only low factor 2 loading ( $-0.148$ – $0.037$ ). The cumulative percentages of this group show a successive increase with water depth from 0.3% at 103 m to 48.2% at 802 m (Fig. 3). Only a slight increase to 66.0% at 1917 m is achieved below. The interval between 901 and 945 m from where on percentages of Assemblage 1 do not increase substantially with depth is close to the upper boundary of the lower MOW core layer in distal settings (Zenk and Armi, 1990). Assemblage 1 from the Gulf of Cadiz contains several species that were previously grouped in the ‘Lower Slope Association’ off southern Portugal (Schönfeld, 1997), e.g. *Cibicides kullenbergi*, *Cribostomoides subglobosum*, *Saccorhiza ramosa* and *Trochammina* spp. There, the first significant increase of ‘Lower Slope Association’ percentages is recognised between 968 and 1103 m, which is only slightly deeper than the comparable change in the Gulf of Cadiz. The major shift to greater proportions of the ‘Lower Slope Association’ occurs off southern Portugal about 1000 m deeper, below the MOW. Although the lower MOW core layer is not very well covered by the present data set from the Gulf of Cadiz, all samples from that depth interval and the one below are dominated by Assemblage 1. They are located close to the margins of the area where the MOW flow impinges the sea floor and hence represent a deep and distal facies (Fig. 4).

Assemblage 2 is characterised by high negative to low positive factor 1 loading ( $-0.599$ – $0.110$ ) and negative to low positive factor 2 loading ( $-0.370$ – $0.062$ ). It may be subdivided into assemblage 2a (factor 1  $<-0.233$ ) and Assemblage 2b (factor 1  $>-1.74$ ) (Fig. 2, Table 1). Assemblage 2a contains typical shelf foraminifera, as *Nonion asterizans* and *Ammonia beccarii*, and species that were previously designated to the ‘Shelf Edge Association’ off southern Portugal (*Amphicoryna scalaris*, *Globobulimina* sp. 324, *Uvigerina elongatastriata*, and *Uvigerina* sp. 221; Schönfeld, 1997). The highest proportions of this subgroup are found in all samples from the shelf and uppermost slope at depths of 103–272 m (Figs. 3 and 4). Between 272 and 496 m, Assemblage 2a is gradually replaced by Assemblage 2b and further de-

Table 1  
Factor loadings and resulting species groups of the living fauna

	Factor 1	Factor 2	Factor 3	Factor 4
Assemblage 1:				
<i>Ammolagena clavata</i>	0.334	−0.083	0.537	0.063
<i>Bulimina striata mexicana</i>	0.509	0.005	−0.085	0.003
<i>Cibicidoides kullenbergi</i>	0.544	0.012	−0.015	0.134
<i>Cibicidoides pseudoungerianus</i>	0.382	0.002	0.056	0.106
<i>Cribrostomoides subglobosum</i>	0.750	0.037	0.092	0.161
<i>Crithionina hispida</i>	0.515	0.005	−0.024	0.021
<i>Cyclammina cancellata</i>	0.308	−0.043	0.061	−0.029
<i>Glomospira charoides</i>	0.549	−0.081	−0.010	0.239
<i>Hyperammina elongata</i>	0.290	−0.032	0.135	−0.062
<i>Hyperammina laevigata</i>	0.519	−0.043	0.000	0.091
<i>Rhizammina algaeformis</i>	0.587	0.008	0.003	0.128
<i>Saccamina sphaerica</i>	0.689	−0.050	0.441	0.187
<i>Saccorhiza ramosa</i>	0.685	−0.067	0.266	0.114
<i>Sphaeroidina bulloides</i>	0.355	−0.063	0.155	0.158
<i>Tolypammina vagans</i>	0.404	0.024	0.349	0.180
<i>Trochammina bellingshauseni</i>	0.260	−0.026	−0.051	−0.031
<i>Trochammina globigeriniformis</i>	0.418	−0.068	0.276	−0.038
<i>Uvigerina mediterranea</i>	0.243	−0.148	0.368	0.082
<i>Uvigerina peregrina parva</i>	0.392	−0.063	0.115	0.018
Assemblage 2a:				
<i>Ammonia beccarii</i>	−0.504	−0.296	−0.754	0.267
<i>Ammoscalaria foliacea</i>	−0.530	−0.317	−0.491	0.220
<i>Amphicoryna scalaris</i>	−0.599	−0.370	−0.473	0.288
<i>Clavulina mexicana</i>	−0.233	0.028	0.292	0.137
<i>Cribrostomoides scitulum</i>	−0.255	−0.313	−0.493	0.225
<i>Globobulimina</i> sp. 324	−0.585	−0.357	−0.839	0.307
<i>Globobulimina turgida</i>	−0.265	−0.164	−0.591	0.222
<i>Hanzawaia rhodiensis</i>	−0.348	−0.170	−0.254	−0.850
<i>Hyalinea balthica</i>	−0.458	−0.289	−0.740	0.278
<i>Nonion asterizans</i>	−0.450	−0.256	−0.640	0.217
<i>Reophax scorpiurus</i>	−0.505	−0.314	0.068	0.185
<i>Reophax subfusiformis</i>	−0.572	−0.271	−0.253	0.267
<i>Uvigerina elongatastriata</i>	−0.355	−0.240	−0.538	0.219
<i>Uvigerina</i> sp. 221	−0.568	−0.364	−0.656	0.266
Assemblage 2b:				
<i>Ammoscalaria tenuimargo</i>	−0.174	−0.157	0.588	0.000
<i>Bigenerina nodosaria</i>	−0.159	−0.147	0.294	0.169
<i>Crithionina mamilla</i>	−0.024	−0.107	0.582	0.004
<i>Gyroidina orbicularis</i>	0.031	0.001	0.113	0.034
<i>Hanzawaia concentrica</i>	0.012	−0.069	0.455	0.067
<i>Hoeglundina elegans</i>	−0.153	−0.087	0.022	−0.972
<i>Jacueella obtusa</i>	−0.133	−0.012	0.490	0.100
<i>Lenticulina rotulata</i>	−0.080	−0.069	−0.013	−0.959
<i>Marsipella elongata</i>	0.107	0.062	0.568	0.147
<i>Melonis barleanum</i>	0.110	−0.182	−0.528	0.264
<i>Nummoloculina contraria</i>	−0.147	−0.058	−0.068	−0.956
<i>Planulina ariminensis</i>	0.035	−0.125	0.480	−0.041
<i>Pyrgo oblonga</i>	−0.129	−0.059	−0.066	−0.947
<i>Reophax bilocularis</i>	−0.172	−0.192	0.299	0.053
<i>Rhabdammina abyssorum</i>	−0.030	−0.169	0.368	0.082
<i>Sigmoilopsis woodi</i>	0.009	−0.064	0.344	0.058
<i>Vulvulina pennatula</i>	0.079	0.000	0.291	0.027

Table 1 (Continued)

	Factor 1	Factor 2	Factor 3	Factor 4
Assemblage 3:				
<i>Ammodiscus tenuis</i>	-0.075	0.930	-0.153	0.033
<i>Cibicides lobatulus</i>	-0.104	0.955	-0.067	0.046
<i>Cibicides refulgens</i>	-0.075	0.932	-0.139	0.035
<i>Cibicoides</i> sp.	-0.037	0.702	0.057	0.171
<i>Deuterammina ochracea</i>	-0.109	0.910	-0.153	-0.155
<i>Discanomalina semipunctata</i>	-0.026	0.830	0.060	0.077
<i>Pyrgoella sphaera</i>	-0.074	0.931	-0.148	0.034
<i>Trifarina angulosa</i>	-0.161	0.329	0.137	0.018
Variance proportion (%)	15.3	12.2	10.1	9.1

creases with depth below. The transition between both subgroups is recorded within that depth interval where the change of ‘Shelf Edge Association’ to ‘Upper Slope Association’ is recognised off southern Portugal (268–498 m). It is still deeper than off NW Africa (100–200 m; Lutze, 1980) or the Bay of Biscay (230–290 m; Pujos-Lamy, 1973).

Assemblage 2b is dominated by species previously assigned to the ‘Upper Slope Association’ off southern Portugal, in particular passive suspension-feeders (*Jacuellia obtusa*, *Marsipella elongata*, *Rhabdammina abyssorum*) (Mackensen, 1987; Linke and Lutze, 1993; Thomsen, 1998) or elevated epibenthic species (*Hanzawaia concentrica*, *Planulina ariminensis*, *Vulvulina pennatula*) that pursue active suspension-feeding (Lutze and Thiel, 1989; Schönfeld, 1997). High proportions of Assemblage 2b are found in samples from the upper MOW core layer and the transition layer above. Most of them are located in distal settings including the top of Faro Drift (Fig. 4). The depth interval where this subgroup dominates the foraminiferal fauna is 396–901 m (Fig. 3).

Species of Assemblage 3 show low, negative factor 1 loading (–0.026 to –0.161) and high, positive factor 2 loading (> 0.329). The majority of species from this group is adapted to epibenthic habitats (*Cibicoides* sp., *Cibicides lobatulus*, *Cibicides refulgens*, *Deuterammina ochracea*, *Discanomalina semipunctata*). Among the Assemblage 3, *Trifarina angulosa* shows a distinctively lower factor 2 loading than the other species (Fig. 2, Table 1). *Trifarina angulosa* is a cosmopolitan,

shallow infaunal species (De Stigter et al., 1998; Schönfeld, 2001), and occurs in low to moderate abundances from subtidal to middle bathyal depths (e.g. Seiler, 1975; Lutze, 1980; Jorissen, 1988). High percentages are recorded from current-swept passages (Hayward et al., 1994), coarse, biogenic sands on the inner shelf (McGann, 1996), and deep, high-energy environments on the outer shelf and upper slope (Mackensen et al., 1985; Violanti, 1996). It is often associated with *C. lobatulus*, *C. refulgens*, and *Planulina exorna* (= *Planulina ariminensis*). Apparently, *T. angulosa* is adapted to strong water turbulences of varying intensity (Mackensen,

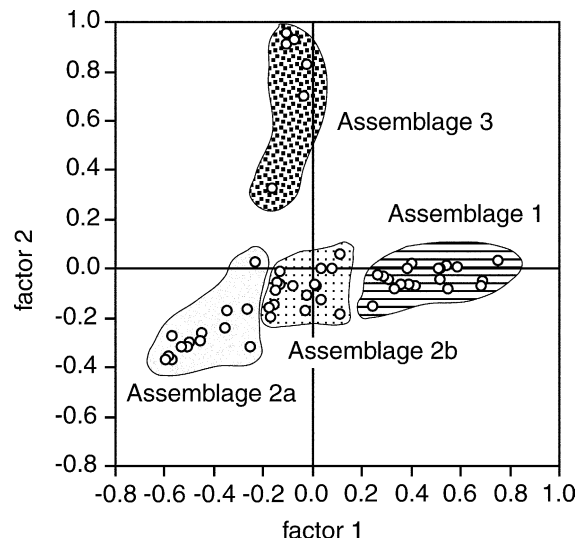


Fig. 2. Orthogonal plot of factor loading from 58 first five ranked species of the living fauna (Table 1).

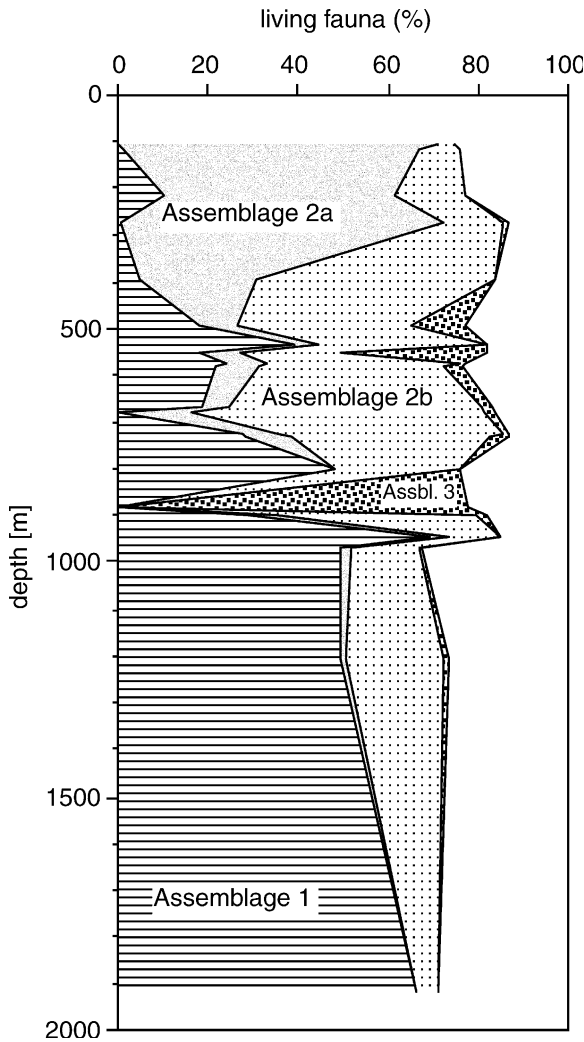


Fig. 3. Depth distribution of cumulative percentages of Assemblages 1, 2a, 2b and 3.

1987). As observations of living specimens attached to large particles have not been reported to date, the species seemingly occupies interstitial microhabitats in the coarse-grained sediments where it may withstand water turbulence. *Trifarina angulosa* is assigned to Assemblage 3 because of their adaptation to similar environments as the other, though epibenthic members of this group.

The highest proportions of Assemblage 3 are found in three samples (M3910-2, M39018-1 and

M39028-3) from 496 to 881 m water depth (Fig. 3). They are located in the immediate flow paths of the upper MOW (Fig. 4). Sample M39010-2 was taken from the centre of a slope-perpendicular channel wherein a current ribbon flows down to the lower MOW core layer (Nelson et al., 1993). The bottom sediment is a coarse sand rich in pebbles and biotritus from bivalves, gastropods, balanids, bryozoans, and deep-water corals ('valley floor facies' after Nelson et al., 1999). A current velocity of  $50 \text{ cm s}^{-1}$  has been recorded 15 m above the seabed 9 km further downstream in that channel (Howe, 1982: Station 21104), whereas  $26 \text{ cm s}^{-1}$  has been measured at an adjacent mooring outside the main flow path (Madelain, 1970: Station C6). Samples M39018-1 and M39028-3 were taken from the centre of the northernmost, slope-parallel channel (locally named 'Fossa Alvarez Cabral'; Faugères et al., 1984). The bottom sediment is a silty fine sand (M39018-1) and medium sand (M39028-3). Flow velocities of up to 40 and  $80 \text{ cm s}^{-1}$  are reported from this channel (Stow et al., 1986), and  $20 \text{ cm s}^{-1}$  from adjacent sites outside the main flow (Madelain, 1970; Howe, 1982). Assemblage 3 therefore characterises proximal settings under the influence of strong and persistent near-bottom currents.

Factor analysis of the dead assemblage only allows the recognition of three species groups, that resemble to a large extent Assemblages 1, 2a and 2b from the living fauna. Index species from Assemblage 3 of the living fauna are grouped together with those of Assemblage 2b. As such, the dead assemblages only display shelf edge conditions, MOW influence, and the environmental conditions of the lower MOW core layer, even though this depth interval is not well represented by the present sample set. The living assemblages trace, however, the top and bottom of the upper MOW core layer and allow separation of environments of high and low hydrodynamic energy.

#### 4.2. Colonisation structures

Samples from locations with a proximal high-energy facies in the eastern Gulf of Cadiz retrieved sand with pebbles, gravel lag and outcrop



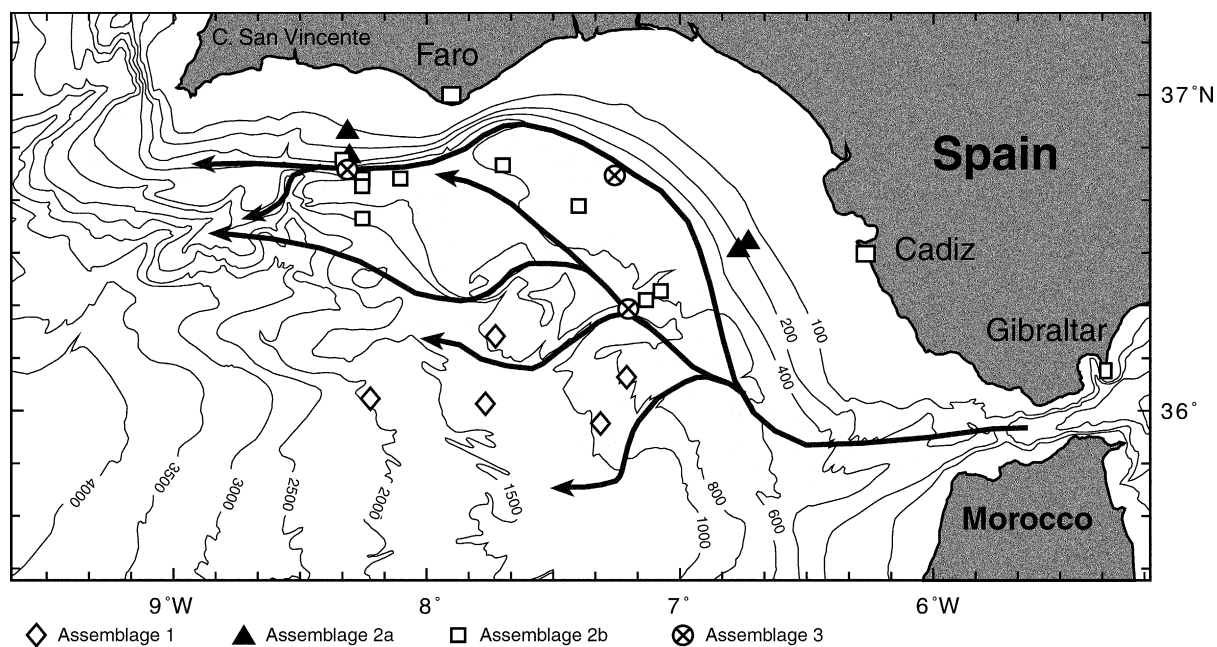


Fig. 4. Location of samples dominated by the foraminiferal assemblages. The area where the immediate MOW undercurrent sweeps along the bottom with salinities  $>36.40$  PSU is marked in light grey (after Heezen and Johnson, 1969; Madelain, 1970; Thorpe, 1976; Ambar and Howe, 1979a,b). Arrows indicate the main MOW flow paths (after Howe, 1982). The bathymetry in metres is given for comparison. Note that the MOW flow may detach from the sea floor at depths below 1200 m.

slabs. Even megacurrent ripples migrating on an omission surface were recovered with a box core on station M39009-1 from a small terrace about 200 m above the valley floor of a slope-perpendicular channel. The standing stock of the entire benthic foraminiferal fauna is with 2.4 living specimens per  $10\text{ cm}^3$  rather low at this location. The epibenthic population density is also low (0.5 specimens per  $10\text{ cm}^2$ ). In the neighbouring sample M39010-2 from the channel centre, attached forms are with 2.0 specimens per  $10\text{ cm}^2$  more frequent than free-living species (1.4 specimens per  $10\text{ cm}^2$ ) but they are not observed on pebbles smaller than 5–7 mm in diameter. Large blocks, however, are densely stocked by epibenthic foraminifers. *Discanomalina semipunctata*, *Cibicides lobatulus*, and *Cibicides refulgens* are most frequent, whereas *Placopsilina confusa*, *Deuterammina ochracea*, *Gavelinopsis translucens*, *Trochammina squammata* and others occur in moderate abundances (Fig. 5, Appendix 4). These species even inhabit ships clinker (sample M39014-1) that is typically avoided elsewhere (Schönfeld,

1997). The blocks are squattered only in the upper part, preferentially above 20 mm (sample M39014-1). Large numbers of foraminifers were found attached to free-standing hydroids collected from the top of an outcrop slab (sample M39010-1). The attachment point of the hydroids is about 140 mm high. Up to 67 foraminifers were found on one hydroid colony (Fig. 5). The average epibenthic population density on the outcrop slabs is with 2.1 specimens per  $10\text{ cm}^2$  very similar to that of the adjacent surface sediment sample M39010-2 (2.0 specimens per  $10\text{ cm}^2$ ) where smaller pebbles are inhabited, preferentially fragments of coral stems. On the flat and shallow outcrop slab (15 mm high) retrieved at the adjacent station M39013-1, where no prominent hydroid colony or other elevated point is available, the average population density is with 0.5 specimens per  $10\text{ cm}^2$  substantially lower. Nonetheless, only large and heavy objects seem to provide a sufficient substrate stability for epibenthic foraminifers in this environment. Sands and small pebbles are permanently reworked, and only specialised endo-

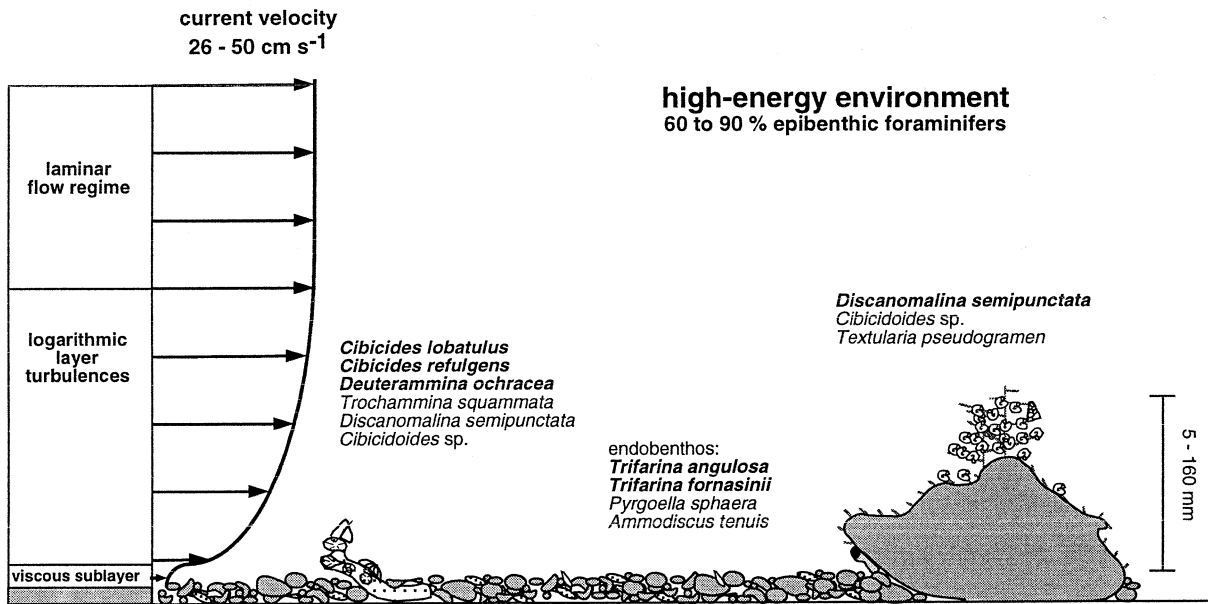


Fig. 5. Epibenthic colonisation structures in high-energy environments in the Gulf of Cadiz (samples M39010-1, M39010-2). A schematic current strength section with distance from the sea bed (not to scale) indicates the different hydrodynamic environments. Frequent species are marked in bold. Epibenthic foraminifers prefer elevated positions on hydroids (grey lines) or large fragments of deep-water corals (stippled).

benthic forms, as for instance *Trifarina angulosa*, could keep pace with erosion and redeposition. Current meter moorings in this area recorded near-bottom velocities ranging from 26 to 50 cm s<sup>-1</sup> (Madelain, 1970; Zenk, 1975b). Epibenthic foraminifers that thrive on suspension-feeding may find even higher food particle fluxes if they settle on more elevated substrates at high current velocities (Muschenheim, 1987; Altenbach et al., 1993; Auffret et al., 1994; Thomsen, 1998). This may explain why the large blocks are only inhabited on the top, and why the population density is lower on shallower objects.

The hydrodynamic environment leaves also an impact in the faunal composition. *Cibicides refulgens* and *Deuterammina ochracea* are frequent in the epibenthic associations in proximal areas. They co-occur with *Quinqueloculina aspera*, *Quinqueloculina intricata* and *Spiroloculina excavata* in the dead assemblage. These species are common in boreal high-energy shelf environments in the Channel area (Calvez, 1958; Rosset-Moulinier, 1986; own data), Irish Sea (Dobson and Haynes, 1973), Rockall Plateau (own data), and off north-

ern Norway (Mackensen et al., 1985; own data), and they were not recorded off south-western Iberia to date.

The situation is quite different in distal settings to the west where the bottom sediment is sandy silt to silty clay (M39020-1 to M39023-3). The standing stock varies between 14 and 39 living specimens per 10 cm<sup>3</sup>. The near-bottom current velocities are in the range of 4–25 cm s<sup>-1</sup> (Heezen and Johnson, 1969; Madelain, 1970; Zenk, 1975b), and hence distinctively lower than in proximal settings. The dominant constituent of the foraminiferal fauna are tubular arenaceous species, mostly *Rhabdammina abyssorum*, that make up to 60% of the living assemblage. The tubes again provide substrates for a diverse epibenthic assemblage of *Crithionina pisum*, *Trochammina squammata*, *Saccammina sphaerica*, *Rosalina anomala* and others (Fig. 6). They also use other objects as colonising substrates, in particular haleciid hydroids, pteropods, mollusc shells, echinoid spines, and sponges. Not all potential objects are colonised, only one of five and one of six in samples M39021-2 and M39022-1 (Ap-

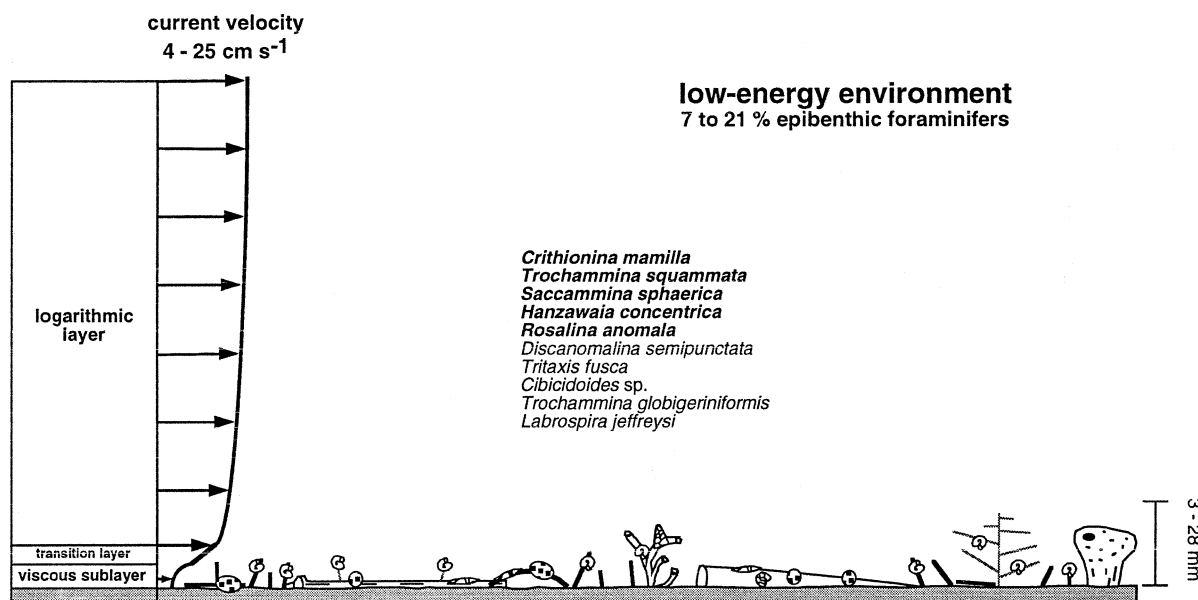


Fig. 6. Epibenthic colonisation structures in low-energy environments (samples M39020-1, M39021-5, M39022-1 and M39023-3). A schematic current strength section with distance from the sea bed (not to scale) indicates the different hydrodynamic environments and the transition layer, where suspended particles are concentrated. Frequent species are marked in bold. Epibenthic foraminifers (stippled: *Crithionina* spp.) use a wide variety of substrates as *Rhabdammina* tubes (thick lines), haleciid hydroids (branched and curved tubes), other hydroids (grey lines), pteropods, mollusc shells, and echinoid spines. They concentrate on a distance of 3–28 mm above the sea bed.

pendix 4). The elevation of the attached foraminifers above the ambient sediment surface typically does not exceed 28 mm, though more elevated substrates are available. This level at about 3 cm height is related to the transition layer between the viscous sublayer close to the sea floor, where a laminar flow regime dominates, and the turbulent logarithmic layer above. The transition layer forms a hydrologic boundary and entraps suspended particles (Altenbach et al., 1988; Rahm and Svensson, 1989). Particle concentration and lateral flux decrease again with distance from the sediment surface and would not provide a substantial higher yield of food particles for epibenthic foraminifers at more elevated positions (Thomsen, 1998).

#### 4.3. Epibenthic assemblages

A compilation of occurrence and settling height above the ambient sediment surface shows that substrates elevated higher than 0.5 cm above the

sea floor are inhabited by epibenthic foraminifera only below 550 m water depth (Fig. 7). There is one exception, sample M39018-1 from the northernmost, slope-parallel channel. In particular *Cibicidoides* sp., *Cibicides lobatulus*, *Discanomalina semipunctata*, *Planulina ariminensis*, *Textularia pseudogramen*, *Trochammina squammata*, and *Vulvulina pennatula* change their settling height between 550 and 576 m. At shallower water depths, they only use low objects even though more elevated substrates are still available. *Cibicides refulgens*, *Cibicides* sp., *Deuterammina ochracea*, *Epistominella rugosa*, *Gavelinopsis translucens*, *Guttulina* sp. (fistulous variety), *Placopsilina confusa*, and *Spiroplectinella sagittula* may also use elevated microhabitats.

The capability of dynamic, elevated microhabitat selection in order to optimise food acquisition under near-bottom currents has been suggested as competitive advantage for selected species forming the 'Epibenthos Group' off southern Portugal (Schönfeld, 1997; Schönfeld and Zahn, 2000).

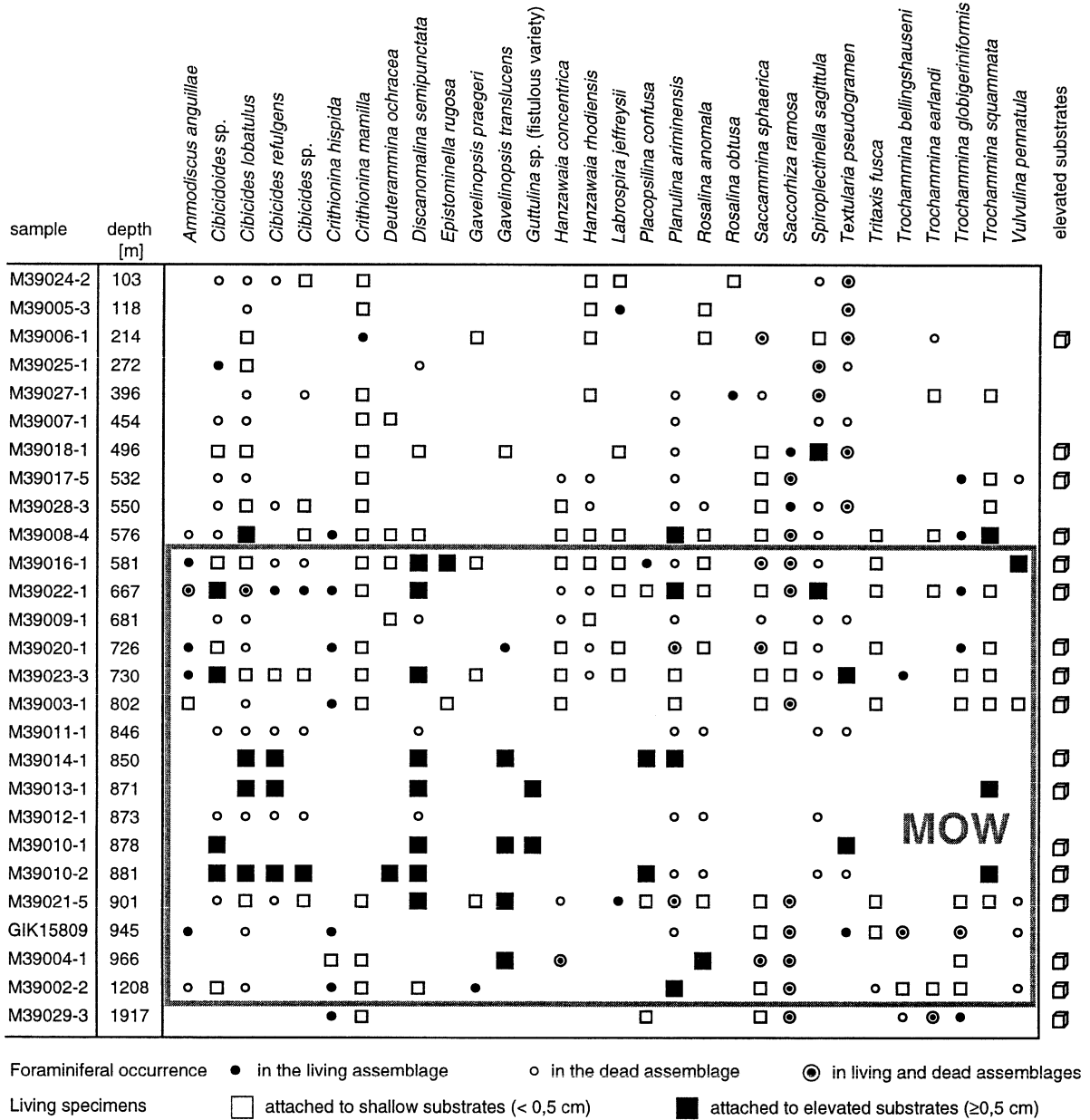


Fig. 7. Depth distribution and attachment elevation of epibenthic foraminifers in the Gulf of Cadiz. Elevated substrates are preferentially occupied in the depth interval of the Mediterranean outflow (grey box).

This association is slightly different in the Gulf of Cadiz area under higher current velocities. With 15 out of 192 species recognised, the elevated epibenthic assemblage comprises 7.8% of the entire faunal spectrum which is more than double their

proportion off southern Portugal (3.6%) where the current strength is lower on average. *Cibicides refulgens*, *Cibicides* sp., *Cibicidoides* sp., *Deuterammima ochracea*, *Discanomalina semipunctata*, *Epistominella rugosa*, *Gavelinopsis translucens*,

*Guttulina* sp. (fistulous variety), *Placopsilina confusa*, *Spiroplectinella sagittula*, *Textularia pseudogramen*, and *Trochammina squammata* supervene to the elevated epibenthic assemblage whereas *Discanomalina coronata* and *Epistominella exigua* were not recorded in the Gulf of Cadiz. *Hanzawaia concentrica* apparently prefers low substrates here even though elevated positions were available in the samples studied.

The majority of species from Assemblage 3, as described above, show elevated microhabitat preferences. Highest proportions of this group were recorded in samples from the immediate MOW flow paths. The depth limit from where below epibenthic species change their settling height and use elevated substrates is in good agreement with the upper boundary of the MOW. Elevated epibenthic foraminifers are therefore considered as reliable indicators for the impact of the MOW contour current in the Gulf of Cadiz, and further downstream off southern Portugal (Schönfeld, 1997).

## 5. Summary and conclusions

The impact of the MOW contour current on recent benthic foraminiferal assemblages is studied in the Gulf of Cadiz, Spain. Factor analyses of living and dead assemblages allows the recognition of four and three different faunal groups. Factor analysis of the dead assemblage displays the shelf edge, upper, and lower MOW environments. The four groups obtained from the living assemblages trace the upper boundaries of MOW core layers, and they discern different biofacies under weak and strong near-bottom currents.

Epibenthic foraminiferal assemblages are dominated by *Discanomalina semipunctata*, *Cibicides lobatulus*, and *Cibicides refulgens* under high near-bottom flow velocities of 26–50 cm s<sup>-1</sup> in the eastern Gulf of Cadiz. The species prefer objects larger than 5–7 mm in diameter that provide a certain microenvironment stability and attach at prominent points. This may be explained by a higher yield of advected food particles with distance from the sediment surface at high current velocities.

In the western Gulf of Cadiz, epibenthic foraminifers such as *Crithionina pisum*, *Trochammina squammata*, *Saccammina sphaerica*, and *Rosalina anomala* use *Rhabdammina abyssorum* tubes and other small objects, but their elevation does not exceed 28 mm. This level is related to the transition layer with high concentrations of suspended particles. Abundance and lateral flux of particles decreases again with distance from the sediment. More elevated positions do therefore not provide a better food yield at low current velocities of 4–25 cm s<sup>-1</sup>.

Colonisation structures, standing stock, and assemblage composition under high versus low current velocities reveals that substrate stability may be a confining environmental variable for benthic foraminifers. Only a few specialised endobenthic species, such as boreal high-energy shelf miliolids and *Trifarina angulosa*, seemingly can withstand permanent winnowing and redeposition. The settling height of epibenthic foraminifera above the ambient sediment is related in proximal and distal settings to the highest lateral flux of suspended food particles within reach from the sea floor. In general, epibenthic foraminifers only use elevated substrates under the influence of near-bottom flow. This pattern may provide a sensitive biotic indicator for the upper depth limit of the MOW current.

The dynamic selection of elevated microhabitats is used as strategy to optimise food acquisition only by a few specialised foraminiferal species. They comprise only 7.8% of all species recognised. Nonetheless this proportion is twice as high as off southern Portugal (3.6%) where the MOW flow is less intense. *Cibicides lobatulus*, *Planulina ariminensis*, and *Vulvulina pennatula* occur in the elevated habitats in both areas. Hence it is justified to assume, that a co-occurrence of these species could serve as indicator of a near-bottom flow regime also in areas outside the MOW flow.

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## Appendix 1. Benthic foraminiferal species which are considered in this paper

Note: Taxonomic references were given by Barker (1960), Ellis and Messina (1940–1978), Schiebel (1992), Timm (1992), and Jones (1994). They are not included in the reference list.

*Ammodiscus anguillae* Höglund 1947

*Ammodiscus tenuis* Brady 1881

*Ammolagena clavata* (Jones and Parker) = *Trochammina irregularis* var. *clavata* Jones and Parker 1860

*Ammonia beccarii* (Linné) = *Nautilus beccarii* Linné 1758

*Ammoscalaria foliacea* (Brady) = *Haplophragmium foliaceum* Brady 1881

*Ammoscalaria tenuimargo* (Brady) = *Haplophragmium tenuimargo* Brady 1882

*Amphicoryna scalaris* (Batsch) = *Nautilus (Orthoceras) scalaris* Batsch 1791

*Bigenerina nodosaria* d'Orbigny 1826

*Bolivina subaenariensis* Cushman 1922

*Bulimina acanthia* Costa 1856

*Bulimina striata mexicana* Cushman = *Bulimina inflata* var. *mexicana* Cushman 1922

*Cibicides lobatulus* (Walker and Jakob) = *Nautilus lobatulus* Walker and Jakob 1798

*Cibicides refulgens* Montfort 1808

*Cibicides* sp.

Note: *Cibicide* sp. differs from *Cibicides lobatulus* by the greater number of chambers in the final coil (8–11), by a distinct umbo on the umbilical side formed by the earlier whorls, and by the nacreous shine of the chamber wall which is recognised in particular in living specimens.

*Cibicoides kullenbergi* (Parker) = *Cibicides kullenbergi* Parker 1953

*Cibicoides pseudoungerianus* (Cushman) = *Truncatulina pseudoungeriana* Cushman 1922

*Cibicoides* sp.

Note: This taxon has been determined as *Cibicoides pseudoungerianus* by Dobson and Haynes (1973), Rosset-Moulinier (1986), and Levy et al. (1995). The tests are planoconvex to slightly biconvex, and the periphery is broadly rounded. It is missing the peripheral carina with circular gutter on the umbilical side, however, as it is typically developed in *Cibicoides pseudoungerianus*.

*Clavulina mexicana* Cushman 1922

*Cribrostomoides scitulum* (Brady) = *Haplophragmium scitulum* Brady 1881

*Cribrostomoides subglobosum* (Sars) = *Lituola subglobosa* Sars 1868

*Crithionina hispida* Flint 1899

*Crithionina mamilla* Goës 1894

*Crithionina pisum* Goës 1896

*Cyclammina cancellata* Brady 1879

*Deuterammina ochracea* (Williamson) = *Rotalina ochracea* Williamson 1858

*Discanomalina coronata* (Parker and Jones) = *Anomalina coronata* Parker and Jones 1857

*Discanomalina semipunctata* (Bailey) = *Rotalina semipunctata* Bailey 1851

*Elphidium crispum* (Linné) = *Nautilus crispus* Linné 1758

*Epistominella exigua* (Brady) = *Pulvinulina exigua* Brady 1884

*Epistominella rugosa* (Phleger and Parker) = *Pseudoparella* (?) *rugosa* Phleger and Parker 1951

*Gavelinopsis praegeri* (Heron-Allen and Earland) = *Discorbina praegeri* Heron-Allen and Earland 1913

*Gavelinopsis translucens* (Phleger and Parker) = *Rotalia translucens* Phleger and Parker 1951

*Globobulimina* sp. 324 Lutze 1980

1995 *Globobulimina auriculata* (Bailey 1851) Levy et al., p. 36, pl. 8, fig. 3.

*Globobulimina turgida* (Bailey) = *Bulimina turgida* Bailey 1851

*Glomospira charoides* (Jones and Parker) = *Trochammina squamata* var. *charoides* Jones and Parker 1860

*Guttulina* sp. (fistulous variety)

Note: The capability of polymorphinid species to anchor in small cavities by using fistulous extensions of their last chamber has been described by Giese (1991) from the Channel off Brittany.

*Gyroidina orbicularis* d'Orbigny 1826

*Hanzawaia concentrica* (Cushman) = *Truncatulina concentrica* Cushman 1918

Note: *Discorbinella bertheloti* (d'Orbigny 1839) of authors.

*Hanzawaia rhodiensis* (Terquem) = *Truncatulina rhodiensis* Terquem 1878

*Hoeglundina elegans* (d'Orbigny) = *Rotalia (Turbinulina) elegans* d'Orbigny 1826

*Hyalinea balthica* (Schröter) = *Nautilus balthicus* Schröter 1783

*Hyperammina elongata* Brady 1884

*Hyperammina laevigata* Wright 1891

*Jacuela obtusa* Brady 1882

*Labrospira jeffreysii* (Williamson) = *Nonionina jeffreysii* Williamson 1858

*Lenticulina rotulata* (Lamarck) = *Cristellaria rotulata* Lamarck 1804

*Marsipella elongata* Norman 1878

*Melonis barleeaanum* (Williamson) = *Nonionina barleeana* Williamson 1858

*Nonion asterizans* (Fichtel and Moll) = *Nautilus asterizans* Fichtel and Moll 1798

*Nummoloculina contraria* (d'Orbigny) = *Biloculina contraria* d'Orbigny 1846

*Placopsilina confusa* Cushman 1920

*Planulina ariminensis* d'Orbigny 1826

*Pyrgo oblonga* (d'Orbigny) = *Biloculina oblonga* d'Orbigny 1839

*Pyrgoella sphaera* d'Orbigny 1839

*Quinqueloculina aspera* d'Orbigny 1826

*Quinqueloculina intricata* Terquem 1878

*Reophax bilocularis* Flint 1899

*Reophax scorpiurus* Montfort 1808

*Reophax subfusiformis* Earland 1933

*Rhabdammina abyssorum* Sars 1868

*Rhizammina algaeformis* Brady 1879

*Rosalina anomala* Terquem 1875

*Rosalina obtusa* d'Orbigny 1846

*Saccamina sphaerica* Sars 1868

*Saccorhiza ramosa* (Brady) = *Hyperammina ramosa* Brady 1879

*Sigmoilopsis woodi* Atkinson 1968

*Sphaeroidina bulloides* d'Orbigny 1826

*Spiroloculina excavata* d'Orbigny 1846

*Spiroplectinella sagittula* d'France 1824

*Textularia pseudogramen* Chapman and Parr 1937

*Tolypammina vagans* Brady 1819

*Trifarina angulosa* (Williamson) = *Uvigerina angulosa* Williamson 1858

*Trifarina fornasinii* (Selli) = *Angulogerina fornasinii* Selli 1948

*Tritaxis fusca* (Williamson) = *Rotalina fusca* Williamson 1858

*Trochammina bellingshauseni* (Brönnimann and Whittaker) = *Globotrochamminopsis bellingshauseni*

Brönnimann and Whittaker 1988

*Trochammina earlandi* (Brönnimann and Whittaker) = *Paratrochammina (Paratrochammina) earlandi*

Brönnimann and Whittaker 1988

*Trochammina globigeriniformis* (Parker and Jones) = *Lituola nautiloidea* var. *globigeriniformis* Parker and Jones 1865

*Trochammina squammata* Jones and Parker 1860

*Uvigerina elongatastriata* (Colom) = *Angulogerina elongatastriata* Colom 1952

*Uvigerina mediterranea* Hofker 1932

*Uvigerina peregrina* Cushman 1923

*Uvigerina peregrina parva* Lutze = *Uvigerina peregrina forma parva* Lutze 1986

Note: *Uvigerina bradyana* (Cushman 1923) of authors.

*Uvigerina* sp. 221 Lutze 1986

1995 *Uvigerina peregrina* (Cushman 1923) Levy et al., p. 38, pl. 8, fig. 11.

*Vulvulina pennatula* (Batsch) = *Nautilus (Orthoceras) pennatula* Batsch 1791

## Appendix 2. Formaminiferal census data of the living assemblage

Only those species are considered which occur in at least one sample among the first five ranked species of the living assemblage.

sample	latitude (*N)	longitude (*W)	depth (m)	<i>Ammoscalaria foliacea</i>	<i>Ammoscalaria tenuimargo</i>	<i>Ammofiscus tenuis</i>	<i>Ammolagena clavata</i>	<i>Ammonia beccarii</i>	<i>Amphicoxyna scalaris</i>	<i>Bigenenerina nocosaria</i>	<i>Bulimina striata mexicana</i>	<i>Cibicides</i> sp.	<i>Cibicides kullenbergi</i>	<i>Cibicides pseudoburgenianus</i>	<i>Cibicides lobatulus</i>	<i>Cibicides refulgens</i>	<i>Clavulina mexicana</i>	<i>Cibrostomoides scitulum</i>	<i>Cibrostomoides subglobosum</i>	<i>Cithionina hispida</i>	<i>Cithionina namilla</i>	<i>Cyclammina cancellata</i>	<i>Deuterammia ochracea</i>	<i>Discanomalina semipunctata</i>	<i>Globobulimina</i> sp. 324	<i>Globobulimina lurida</i>	<i>Glomospira charoites</i>	<i>Gyroidina orbicularis</i>	<i>Hanzawaia concentrica</i>	<i>Hanzawaia modiensis</i>	<i>Hoeglundina elegans</i>	<i>Hyalina bathica</i>	
M39024-2	36.882	8.313	103	1.7	-	-	-	3.8	6.4	2.3	-	-	-	-	-	-	1.2	-	-	0.6	-	-	-	13.1	3.8	0.3	-	-	5.0	-	0.3		
M39005-3	36.535	6.736	118	8.1	-	-	-	1.2	10.5	2.4	-	-	-	-	-	-	6.5	-	-	-	-	-	-	-	10.1	3.2	0.4	-	-	2.5	-	1.2	
M39006-1	36.512	6.774	214	0.8	-	-	-	0.4	3.2	9.2	-	-	-	-	-	-	0.8	2.0	-	-	0.4	0.8	-	-	8.0	1.2	0.4	-	-	2.4	0.4	-	
M39025-1	36.803	8.312	272	0.9	-	-	-	3.6	0.9	-	-	-	-	-	-	-	-	0.9	-	-	-	-	-	-	12.5	28.6	-	-	-	-	0.9	-	
M39027-1	36.782	8.317	396	4.2	4.2	-	0.8	-	-	0.4	3.8	0.4	-	-	-	-	0.4	0.8	-	-	0.8	-	-	-	1.3	-	0.4	-	-	0.4	0.8	-	
M39018-1	36.753	7.252	496	-	-	-	3.8	-	-	5.9	-	2.7	-	0.8	2.1	-	0.8	0.8	2.1	-	1.3	0.8	-	4.4	-	0.4	0.8	-	-	-	0.8	-	
M39017-5	36.650	7.410	532	-	3.4	-	18.9	-	-	15.5	-	-	-	-	-	-	-	-	-	2.1	-	-	-	-	-	-	-	-	-	-	-	-	
M39028-3	36.771	8.318	550	1.9	1.9	-	7.5	-	-	2.8	-	-	-	-	2.4	-	0.9	-	-	-	7.1	0.9	-	-	-	-	-	-	0.9	-	0.9	-	
M39008-4	36.381	7.076	576	-	6.5	-	2.1	-	3.6	3.3	-	-	-	0.9	0.6	-	0.3	-	0.3	1.2	3.3	0.3	0.3	0.1	0.3	0.6	0.9	-	1.8	0.6	0.9	-	
M39016-1	36.779	7.706	581	-	2.7	-	-	2.0	4.0	3.6	-	1.8	-	-	1.3	-	1.3	0.4	-	-	1.3	1.3	0.2	0.9	-	0.4	-	-	0.7	0.9	-	-	
M39022-1	36.712	8.260	667	-	-	-	2.0	-	1.6	2.4	-	0.5	-	0.4	0.4	0.4	0.8	1.2	1.2	4.2	1.2	-	0.8	-	-	1.2	0.4	-	-	-	-	-	
M39009-1	36.350	7.142	681	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2.0	-	-	-	-	-	-	-	16.3	16.3	-	-
M39020-1	36.738	8.106	726	-	1.2	-	2.9	-	1.2	3.5	-	0.9	-	-	-	-	1.2	1.2	0.6	2.1	0.9	-	-	-	-	-	0.6	-	1.2	-	-	-	
M39023-3	36.736	8.254	730	-	2.8	-	14.3	-	-	3.5	-	0.9	-	-	0.9	0.7	-	0.7	2.1	-	5.2	2.1	-	0.8	-	-	-	0.1	-	-	-	-	
M39003-1	36.111	7.223	802	-	-	-	5.1	-	-	2.6	-	-	-	-	-	-	-	0.9	0.9	1.5	4.3	-	-	-	-	-	0.4	-	-	-	-	-	
M39010-2	36.321	7.208	881	-	-	10.0	-	-	-	-	2.5	-	-	-	17.5	12.5	-	-	-	-	-	-	10.0	5.0	-	-	-	-	-	-	-	-	-
M39021-5	36.608	8.255	901	-	6.6	-	9.2	-	-	1.3	-	-	-	-	-	-	1.3	-	-	-	10.2	2.6	-	1.3	-	-	-	-	-	-	-	-	-
GIK15809	35.962	7.314	945	-	-	-	3.2	-	-	0.8	-	-	-	-	-	-	3.8	1.5	1.0	-	20.6	-	-	-	-	-	-	-	-	-	-	-	-
M39004-1	36.237	7.732	966	-	-	-	7.6	-	-	1.0	-	-	-	-	-	-	0.3	1.0	4.8	2.9	-	-	-	-	-	1.0	2.4	-	0.3	-	-	-	-
M39002-2	36.027	7.775	1208	-	-	-	7.6	-	-	0.8	2.3	0.8	-	-	-	-	1.5	3.0	12.9	0.2	2.3	-	0.8	-	-	-	-	-	-	-	-	-	-
M39029-3	36.041	8.233	1517	-	-	-	3.9	-	-	-	-	-	6.7	-	-	-	-	7.7	1.0	0.2	1.0	-	-	-	-	-	-	-	-	-	-	-	-

sample	<i>Hyperammia elongata</i>	<i>Hyperammia laevigata</i>	<i>Jaculella obtusa</i>	<i>Lenticulina rotulata</i>	<i>Marsipella elongata</i>	<i>Melonis barthelemyi</i>	<i>Nonion astenzans</i>	<i>Nummoculina contraria</i>	<i>Planulina aminensis</i>	<i>Pygoella sphaera</i>	<i>Pyrgo oblonga</i>	<i>Reophax biculans</i>	<i>Reophax scorpiurus</i>	<i>Reophax subuliformis</i>	<i>Rhabdammina abyssorum</i>	<i>Rhizammina algaeformis</i>	<i>Saccamina sphaerica</i>	<i>Saccorhiza ramosa</i>	<i>Stigmolopsis woodi</i>	<i>Sphaerotina bulloides</i>	<i>Tolypammia vagans</i>	<i>Trifarina angulosa</i>	<i>Trochammina bellingshauseni</i>	<i>Trochammina globiferiformis</i>	<i>Uvigerina elongatastrata</i>	<i>Uvigerina mediterranea</i>	<i>Uvigerina peregrina parva</i>	<i>Uvigerina</i> sp. 221	<i>Vulvulina pennatula</i>	counted specim.	standing stock (#/10 cm <sup>3</sup> )
M39024-2	-	-	-	-	-	0.3	10.2	-	-	-	0.6	5.2	9.0	-	-	-	-	-	-	-	-	-	-	1.7	-	-	9.3	-	343	30.8	
M39005-3	-	-	-	-	-	2.8	1.2	-	-	-	3.6	4.4	4.0	-	-	-	-	-	-	-	-	-	-	4.9	0.8	-	8.1	-	247	45.2	
M39006-1	-	0.4	-	-	-	2.4	-	-	-	-	1.6	0.4	2.9	-	0.4	-	-	-	0.4	-	-	-	-	21.3	8.4	8.8	-	249	46.3		
M39025-1	-	-	-	-	-	9.8	5.4	-	-	-	0.9	0.9	0.9	2.7	-	-	-	-	-	-	-	-	-	14.3	0.9	-	1.8	-	112	12.7	
M39027-1	-	-	0.4	0.4	1.7	2.1	-	-	-	-	8.0	6.8	1.3	34.2	-	-	-	-	-	0.4	-	-	-	5.1	3.4	-	1.3	-	237	53.8	
M39018-1	-	-	-	0.8	24.9	1.7	-	-	-	-	0.8	2.5	3.3	2.3	-	3.8	2.7	-	-	0.8	2.9	-	-	0.4	2.5	-	-	-	239	20.8	
M39017-5	-	-	-	-	12.2	-	-	-	-	-	1.7	2.6	2.6	-	-	6.2	9.9	2.6	-	1.7	-	-	0.9	-	1.7	-	-	-	117	14.6	
M39028-3	-	2.8	-	-	-	-	-	-	-	-	1.9	1.9	4.7	3.8	-	0.9	1.9	-	-	0.9	30.2	-	-	-	5.7	-	-	-	106	22.4	
M39008-4	0.6	0.6	3.3	-	13.2	0.3	-	0.7	-	-	0.6	2.4	9.2	-	0.1	7.8	-	-	1.2	0.3	-	-	0.3	-	8.0	-	0.9	-	338	21.7	
M39016-1	-	0.4	5.6	-	18.0	0.9	-	-	-	-	0.9	0.9	1.3	6.9	-	2.2	1.3	0.4	-	6.7	-	-	-	8.9	-	-	0.2	225	17.5		
M39022-1	0.4	-	2.4	0.4	8.3	0.8	-	0.4	0.4	-	1.6	0.4	36.7	-	2.8	5.2	-	-	-	-	-	-	0.8	-	2.4	-	1.6	252	32.9		
M39009-1	-	-	-	16.3	-	-	-	16.3	-	16.3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	6	2.4		
M39020-1	-	-	1.5	0.9	9.7	0.3	-	0.9	-	-	2.9	2.4	1.2	26.0	-	7.1	4.1	0.3	0.9	1.5	-	-	0.3	0.6	7.7	0.3	0.9	-	339	39.0	
M39023-3	-	-	1.4	1.4	12.0	-	-	0.1	-	-	0.7	7.1	2.8	15.8	-	2.9	0.8	0.7	0.7	1.5	0.7	0.7	0.8	-	8.0	-	-	141	36.9		
M39003-1	-	1.7	-	0.9	10.4	0.9	-	0.4	-	-	1.7	-	7.7	1.7	1.5	3.2	-	-	-	-	-	-	6.0	-	20.5	2.6	0.2	117	14.2		
M39010-2	-	-	-	-	-	-	-	-	10.0	-	-	-	-	-	-	-	-	-	-	-	-	10.0	-	-	-	-	-	-	10	3.5	
M39021-5	5.3	-	-	1.3	-	-	-	1.3	-	-	4.0	5.3	-	15.5	-	4.0	3.0	-	-	-	-	-	2.0	-	2.6	-	-	-	76	13.5	
GIK15809	-	-	-	3.0	1.5	-	-	-	-	-	-	-	-	4.6	-	1.1	17.5	-	-	1.0	-	13.0	0.8	-	9.1	-	-	131	9.2		
M39004-1	1.4	7.1	-	2.0	-	8.3	-	-	-	-	-	-	1.4	-	7.1	4.9	-	-	0.4	0.3	-	-	0.1	1.0	7.4	1.7	-	296	40.7		
M39002-2	7.0	-	-	1.5	2.5	3.0	-	0.8	-	-	-	-	12.7	0.8	2.5	3.0	-	-	-	-	1.5	-	1.5	3.2	-	1.5	-	132	19.6		
M39029-3	-	1.0	-	-	1.9	-	-	-	-	-	1.0	1.0	-	1.0	12.5	7.2	15.4	-	-	1.0	5.8	-	1.0	-	-	-	-	104	15.7		



### Appendix 3. Foraminiferal census data of the dead assemblage

Only those species are considered which occur in at least one sample among the first five ranked species of the dead assemblage.

sample	latitude (°N)	longitude (°W)	depth (m)	<i>Ammoscalaria foliacea</i>	<i>Ammolagena clavata</i>	<i>Ammonia beccarii</i>	<i>Amphicoryna scalaris</i>	<i>Bigenerina notosaria</i>	<i>Bolivina subaenariensis</i>	<i>Bullimina acanthia</i>	<i>Bullimina striata mexicana</i>	<i>Cibicides</i> sp.	<i>Cibicides kullenbergi</i>	<i>Cibicides pseudoungeriensis</i>	<i>Cibicides lobatulus</i>	<i>Cibicides refulgens</i>	<i>Clavulina mexicana</i>	<i>Cribrostomoides scitulum</i>	<i>Cribrostomoides subglobosum</i>	<i>Cyclammmina cancellata</i>	<i>Discanomalina semipunctata</i>	<i>Elphidium crispum</i>	<i>Globobulimina</i> sp. 324	<i>Globobulimina turrida</i>	<i>Glomospira charoides</i>	<i>Gyroldina orbicularis</i>	<i>Hanzawaia concentrica</i>	<i>Hanzawaia rhodiensis</i>	<i>Hoeglundina elegans</i>	<i>Hyalinea bathica</i>	<i>Hyperammmina elongata</i>					
M39024-2	36.882	8.313	103	0.7	-	7.4	6.8	3.5	-	-	-	1.8	-	-	2.1	1.4	-	-	-	-	-	1.8	3.2	0.4	-	-	-	-	2.5	-	6.7	-				
M39005-3	36.535	6.736	118	0.4	-	12.9	10.3	3.4	0.4	-	-	-	-	-	4.6	-	-	-	-	-	-	0.4	8.7	1.5	-	-	-	-	1.9	-	13.7	-				
M39006-1	36.512	6.774	214	0.3	-	14.4	4.5	2.5	5.1	1.4	-	-	-	2.8	2.0	-	-	-	-	-	-	0.4	2.5	0.3	-	-	-	-	-	-	27.7	-				
M39025-1	36.803	8.312	272	-	-	0.7	6.4	0.7	-	-	-	-	-	4.3	-	-	1.4	-	-	-	-	0.7	1.4	7.1	-	-	-	-	-	3.5	5.0	-				
M39027-1	36.782	8.317	396	-	-	-	7.2	4.6	-	-	1.5	-	-	7.7	5.7	-	1.0	-	-	-	-	-	1.0	9.3	-	-	-	2.1	-	2.1	3.1	-				
M39007-1	36.455	6.916	454	-	-	7.7	0.8	1.9	-	3.4	-	1.9	-	3.1	3.1	-	0.8	-	-	-	-	2.7	-	1.9	-	-	-	-	-	5.7	10.0	-				
M39018-1	36.753	7.252	496	-	-	-	11.0	6.7	-	-	0.6	6.1	-	3.1	5.5	-	0.6	-	-	-	-	-	0.6	-	-	-	-	-	-	-	1.8	7.4	-			
M39017-5	36.650	7.410	532	-	0.5	-	0.5	22.5	-	-	1.4	0.5	0.9	3.8	2.3	-	4.7	-	-	-	-	-	-	-	-	-	-	-	-	-	5.8	8.0	-			
M39028-3	36.774	8.319	550	-	-	-	3.5	3.9	-	-	1.3	1.8	3.1	0.4	5.7	0.4	0.9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2.6	1.3	4.4	2.6	
M39008-4	36.381	7.076	576	-	-	-	9.8	7.6	-	-	2.2	-	-	2.2	5.1	-	6.2	0.4	1.1	1.1	2.9	-	-	0.4	0.7	0.7	0.7	-	-	-	-	0.7	-	0.7	-	
M39016-1	36.779	7.706	581	-	0.2	-	9.8	16.7	-	-	0.6	5.1	-	3.2	2.8	0.2	3.0	0.2	0.4	0.4	-	-	-	0.4	-	0.2	0.6	0.2	0.6	0.6	-	-	0.6	-		
M39022-1	36.712	8.260	667	-	0.8	-	4.9	8.7	-	-	1.9	1.9	-	3.8	2.3	-	3.4	0.4	2.6	0.4	1.1	-	-	-	-	-	1.1	0.4	0.8	0.8	-	-	0.4	-		
M39009-1	36.350	7.142	681	-	-	0.5	0.5	1.5	-	6.7	1.0	7.2	-	5.1	2.1	-	1.0	-	-	-	-	1.0	0.5	-	-	-	9.2	-	8.2	-	-	-	-	-		
M39020-1	36.738	8.106	726	-	-	0.5	0.5	6.0	1.1	-	-	-	-	1.6	6.6	-	1.6	-	-	-	-	0.5	-	0.5	-	-	2.7	3.8	4.9	6.6	-	-	-	-		
M39023-3	36.736	8.254	730	-	-	-	3.4	7.2	-	0.3	6.8	3.4	-	0.3	6.1	-	3.4	0.3	-	-	-	-	1.0	0.3	-	-	0.3	0.3	1.0	4.4	-	-	-	-		
M39003-1	36.111	7.223	802	-	1.5	-	0.5	3.5	-	-	7.4	-	-	1.5	0.5	-	3.0	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2.5	0.5	
M39011-1	36.271	7.215	846	-	-	2.8	-	0.4	-	0.7	0.4	6.7	-	1.4	30.7	2.5	-	-	-	-	-	3.5	11.3	-	0.4	-	-	-	-	-	-	-	-	1.4	1.8	
M39012-1	36.245	7.220	873	-	-	5.6	-	0.9	-	0.9	0.5	8.0	-	2.3	15.5	0.9	0.5	-	-	-	-	6.6	12.7	-	-	-	-	-	-	-	-	-	-	1.9	1.9	
M39010-2	36.321	7.208	881	-	-	8.8	-	0.9	-	-	6.1	-	-	3.5	15.8	0.9	-	-	-	-	-	16.7	7.9	-	-	-	-	-	-	-	-	-	-	0.9	-	
M39021-5	36.608	8.255	901	-	2.9	-	0.9	5.4	-	0.9	1.1	6.0	0.6	6.0	2.0	2.3	0.9	-	-	-	-	1.7	0.9	-	-	-	0.3	0.3	-	-	-	-	-	0.6	-	
GIK15809	35.952	7.314	945	-	-	-	1.8	-	-	-	8.7	-	-	3.8	5.0	-	1.1	-	-	-	-	-	-	-	-	2.2	-	-	-	-	-	-	-	-	-	
M39004-1	36.237	7.732	966	-	-	2.4	-	4.4	-	-	-	-	-	9.8	5.0	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
M39002-2	36.027	7.775	1208	-	-	5.2	-	1.3	1.3	-	9.4	-	-	0.4	0.4	-	-	-	-	-	-	-	-	-	2.4	-	3.3	-	-	-	-	-	-	-	0.8	-
M39029-3	36.041	8.233	1917	-	6.3	-	-	-	-	-	3.6	-	11.7	-	-	-	-	-	-	-	-	-	-	-	5.8	0.4	-	-	-	-	-	-	-	-	3.0	-

sample	<i>Hyperammmina laevigata</i>	<i>Jacuelia obtusa</i>	<i>Lenticulina rotulata</i>	<i>Marsipella elongata</i>	<i>Melonis barleanum</i>	<i>Nonion astersans</i>	<i>Nummuloculina conizaria</i>	<i>Planulina ariminensis</i>	<i>Pyrgoella sphaera</i>	<i>Pyrgo oblonga</i>	<i>Reophax bilocularis</i>	<i>Reophax scorpionus</i>	<i>Reophax subuliformis</i>	<i>Rhabdammina abyssorum</i>	<i>Rhizammina algaeformis</i>	<i>Saccarina sphaerica</i>	<i>Saccorhiza ramosa</i>	<i>Sigmaliopsis woodi</i>	<i>Sphaeroidina bulloides</i>	<i>Tolypammmina vagans</i>	<i>Trifarina angulosa</i>	<i>Trochammmina bellingshauseni</i>	<i>Trochammmina globigeriniformis</i>	<i>Uvigerina elongatistriata</i>	<i>Uvigerina mediterranea</i>	<i>Uvigerina peregrina parva</i>	<i>Uvigerina peregrina</i>	<i>Uvigerina</i> sp. 221	<i>Vulvulina pennatula</i>	counted specim.	abundance (#/10 cm <sup>3</sup> )			
M39024-2	-	-	-	-	1.1	27.4	-	-	-	-	0.4	0.4	1.1	-	-	-	-	-	0.4	-	-	-	-	-	-	-	-	-	-	12.3	285	982.0		
M39005-3	-	-	-	-	1.9	6.5	-	-	-	-	-	2.7	3.0	0.8	-	-	-	-	0.8	0.4	-	-	-	0.4	1.5	-	-	-	-	6.5	263	192.3		
M39006-1	-	0.3	-	-	2.5	0.6	-	-	-	-	-	0.3	-	0.3	-	0.3	-	0.3	0.8	0.8	-	-	-	0.8	7.6	-	-	9.0	354	2087.6				
M39025-1	-	-	-	-	1.4	-	-	-	-	-	1.4	-	-	-	-	-	-	-	2.8	-	-	-	-	13.5	7.8	-	-	29.1	141	2281.2	-	-	-	
M39027-1	-	-	0.5	-	2.6	-	-	1.0	-	-	0.5	0.5	-	0.5	-	0.5	-	-	5.2	-	-	-	-	3.1	8.2	-	-	11.3	194	11709.3	-	-	-	
M39007-1	-	-	0.4	-	9.6	0.8	-	1.9	-	1.1	-	-	-	-	-	-	-	-	0.4	-	-	-	-	-	8.4	-	-	7.7	261	15817.8	-	-	-	
M39018-1	-	0.6	1.2	-	0.6	5.5	-	1.8	-	-	-	-	1.2	0.6	-	-	-	1.8	7.9	-	-	-	-	0.6	16.6	-	-	6.1	163	3795.8	-	-	-	
M39017-5	-	-	-	-	0.5	3.3	-	2.3	-	-	0.5	-	-	-	-	-	-	4.2	-	-	-	-	-	-	28.6	-	-	2.8	0.5	213	3462.7	-	-	-
M39028-3	-	-	1.3	0.4	7.0	-	-	4.4	-	-	-	-	-	0.4	-	-	-	1.8	7.9	-	-	-	-	1.3	19.7	-	-	3.9	228	22287.4	-	-	-	
M39008-4	-	0.4	6.9	0.7	0.7	-	-	5.1	0.4	0.4	-	0.4	1.5	4.0	-	0.7	2.2	4.7	-	-	-	-	-	-	4.4	-	-	1.5	-	275	533.9	-	-	-
M39016-1	-	0.6	5.3	0.4	1.9	-	-	5.8	-	0.2	-	-	0.2	2.1	0.4	0.4	2.6	0.4	0.2	-	-	-	-	0.4	11.8	-	-	2.1	468	674.4	-	-	-	
M39022-1	-	0.4	1.1	2.3	1.5	-	-	7.9	-	-	-	0.4	-	15.8	2.3	5.7	1.5	-	0.8	-	-	-	-	-	7.5	-	-	2.3	-	265	498.8	-	-	-
M39009-1	-	-	1.0	-	9.2	-	0.5	6.7	-	1.0	-	-	-	-	0.5	-	-	-	-	-	-	-	-	-	10.1	-	-	4.1	-	195	11935.9	-	-	-
M39020-1	-	-	0.5	1.1	2.7	0.5	-	3.8	-	-	-	0.5	1.6	2.7	3.3	1.6	3.3	1.1	-	-	-	-	-	-	12.1	-	-	4.9	-	182	1655.2	-	-	-
M39023-3	-	-	1.7	0.3	1.0	1.0	-	6.8	-	-	-	0.7	-	0.3	0.3	1.7	2.0	-	-	-	-	-	-	0.3	16.0	1.0	-	2.7	-	293	9616.0	-	-	-
M39003-1	-	-	1.5	0.5	2.0	-	-	4.5	-	-	-	-	-	0.5	0.5	-	-	-	-	-	-	-	-	-	37.6	4.0	-	-	-	202	848.6	-	-	-
M39011-1	-	-	0.7	-	3.9	-	0.4	1.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0.4	-	-	0.4	-	283	808.1	-	-	-
M39012-1	-	-	0.5	-	2.8	0.5	-	2.3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2.3	-	-	0.5	-	213	2600.9	-	-	-
M39010-2	-	-	0.9	-	0.9	-	-	0.9	-	-	-	-	-	-	-	-	-	0.9	-	-	-	-	-	-	-	-	-	-	-	114	2403.0	-	-	-
M39021-5	-	-	1.7	0.3	2.0	0.3	-	4.9	-	-	-	0.9	4.9	-	1.1	0.9	2.3	0.6	-	-	-	-	0.3	-	5.1	2.6	-	6.6	0.6	350	3123.3	-	-	-
GIK15809	-	-	2.7	-	4.4	-	-	2.7	-	0.5	-	-	0.7	-	0.1	0.7	-	-	-	-	-	-	0.5	0.5	23.4	5.4	-	-	6.5	184	825.9	-	-	-
M39004-1	2.4	-	3.3	-	9.8	-	-	-	-	0.8	-	-	-	-	29.3	13.0	-	-	-	-	-	-	-	-	6.5	2.4	-	-	-	123	75.3	-	-	-
M39002-2	-	-	1.7	0.9	2.6	-	-	0.4	-	-	-	-	-	9.4	0.9	3.0	3.0	-	-	-	-	-	0.4	-	4.3	6.4	-	-	1.7	233	34.7	-	-	-
M39029-3	-	-	-	3.6	-	-	-	0.9	-	0.4	0.9	-	2.2	-	0.9	8.1	-	-	-	-	-	-	-	-	-	-	-	-	-	223	67.5	-	-	-

#### Appendix 4. Recordings of the living epibenthic foraminiferal species

Once the height above the ambient sediment is recognised, the maximum attachment elevation in mm is added. Abbreviation of substrate types: br, bryozoans; co, coral fragments; cr, crinoids; ech, echinoid spines; hy, hydroids; mo, mollusk fragments; pch, polychaet tubes; pt, pteropods; py, pyrite nodules; qz, quartz; sc, scaphopods; sk, ship's clinker; sp, sponges; taf, tubular arenaceous foraminifers. \*, settling on other epizoans (mostly hydroids) on the large objects, attachment elevation is given in mm above the base of the host, and the squatter density refers to one single colony. §, specimens squatter small cavities in the slab surface. Note that the smaller surface area in the size fraction 250–2000 µm is due to splits been made to reduce the sample to a practical size.

sample	latitude (°N)	longitude (°W)	depth (m)	grainsize fractions, no. of other objects	substrate types	<i>Ammodiscus anguillae</i>	<i>Cibicides</i> sp.	<i>Cibicides lobatulus</i>	<i>Cibicides refugens</i>	<i>Cibicides</i> sp.	<i>Crithionina hispida</i>	<i>Crithionina mamilla</i>	<i>Deuterammina ochracea</i>	<i>Discanomalina semipunctata</i>	<i>Epistominella rugosa</i>	<i>Gavelinopsis praegeri</i>	<i>Gavelinopsis translucens</i>	<i>Gutulina</i> sp. (fistulous variety)	<i>Hanzawaia concentrica</i>
M39024-2	36.882	8.313	103	250-2000 µm >2000 µm	mo, qz, pch mo	-	-	-	1	-	1	-	-	-	-	-	-	-	-
M39005-3	36.535	6.736	118	250-2000 µm >2000 µm	taf, mo mo, pch	-	-	-	-	-	1	-	-	-	-	-	-	-	-
M39006-1	36.512	6.774	214	250-2000 µm >2000 µm	mo pch 2 hy	-	-	2	-	-	-	-	-	-	2	-	-	-	-
M39027-1	36.782	8.317	396	250-2000 µm	mo, taf	-	-	-	-	-	2	-	-	-	-	-	-	-	-
M39007-1	36.455	6.916	454	>2000 µm	mo	-	-	-	-	-	1	2	-	-	-	-	-	-	-
M39018-1	36.753	7.252	496	250-2000 µm >2000 µm	mo, hy, taf hy, br, taf, pt, mo	-	3	4	-	-	-	-	5	-	-	-	-	-	-
M39017-5	36.650	7.410	532	250-2000 µm >2000 µm	mo, taf pt pch	-	7/3	2	-	-	4	-	11/3	-	-	4	-	-	-
M39028-3	36.771	8.318	550	250-2000 µm >2000 µm	mo, pt mo	-	-	2	1	-	7	-	-	-	-	-	-	-	1
M39008-4	36.381	7.076	576	250-2000 µm >2000 µm	pt, mo, taf pt, mo, taf, br co, taf, cr, sc	-	4	2	-	-	6	1	-	-	-	-	-	-	5
M39016-1	36.779	7.706	581	250-2000 µm >2000 µm	mo, hy, taf, br mo, pt, taf, br, hy hy, taf, pch	-	4	-	-	-	10	-	1	-	-	-	-	-	2
M39022-1	36.712	8.260	667	250-2000 µm >2000 µm	taf, pt, br taf, hy, pt	-	3/4	-	1	-	9	-	2	-	-	-	-	-	-
M39009-1	36.350	7.142	681	250-2000 µm >2000 µm	mo mo	-	1/23	-	-	-	2/2	-	26/28	-	-	-	-	-	-
M39020-1	36.738	8.106	726	250-2000 µm >2000 µm	taf, mo, pt mo, taf taf, sp	-	-	-	-	-	3	-	-	-	-	-	-	-	1
M39023-3	36.736	8.254	730	250-2000 µm >2000 µm	mo, taf mo, pt, taf, hy, br ech, sk, sp	-	1	-	1	-	4	-	-	-	-	-	-	-	2
M39003-1	36.111	7.223	802	250-2000 µm >2000 µm	pt, mo pt, taf, mo komokiacea	1	-	-	-	-	1	-	-	-	-	-	-	-	2
M39014-1	36.272	7.215	850	1 ship's clinker	sp, slag	-	17/40	10/30	-	-	-	-	4/20	-	-	-	3/40	-	-
M39013-1	36.320	7.207	871	1 outcrop slab	hy, sp, sandstone	-	5/15	7/15	-	-	-	-	6/15	-	-	-	-	-	3/15 <sup>§</sup>
M39010-1	36.321	7.208	878	3 outcrop slabs	hy*, sp*	-	2/9	-	-	-	-	-	83/3-18	-	-	1	1	-	-
M39010-2	36.321	7.208	881	>2000 µm	co, mo, ech	-	1/6	7/10	5/5	2/5	-	4/6	2/5	-	-	-	-	-	-
M39021-5	36.608	8.255	901	250-2000 µm >2000 µm	mo, pt py, pt, mo, hy 5 (1 colonized)	-	-	3	-	1	3	-	4/11	-	1	1/6	-	-	-
G1K15809	35.962	7.314	945	250-1000 µm	pt	-	-	1	-	-	1	-	-	-	-	-	-	-	-
M39004-1	36.237	7.732	966	1000-2000 µm	taf	-	-	-	-	-	5	7	-	-	-	-	-	-	-
M39002-2	36.027	7.775	1208	250-2000 µm >2000 µm	pt, taf, pch pt, hy, taf hy, pt, mo sp, taf	-	-	-	-	-	10	6	-	-	-	-	1/7	-	-
M39029-3	36.041	8.233	1917	250-2000 µm >2000 µm	mo pt, sk	-	1	-	-	-	-	-	1	-	-	-	-	-	-

sample	grainsize fractions, no. of other objects														total number of specimens	examined surface area (cm <sup>2</sup> )	average population density (#/10 cm <sup>2</sup> )	maximum squattering density (#/object)			
		<i>Hanzawaia rhodiensis</i>	<i>Labrospira jeffreysii</i>	<i>Piacopsilina confusa</i>	<i>Planulina ariminensis</i>	<i>Rosalina anomala</i>	<i>Rosalina obtusa</i>	<i>Saccammina sphaerica</i>	<i>Saccorniza ramosa</i>	<i>Spiroplectinella sagittula</i>	<i>Textularia pseudogramen</i>	<i>Triaxis fusca</i>	<i>Trochammina bellingshauseni</i>	<i>Trochammina earlandi</i>					<i>Trochammina globigeriniformis</i>	<i>Trochammina squammata</i>	<i>Vulvulina pennatula</i>
M39024-2	250-2000 µm	10	1	-	-	-	1	-	-	-	-	-	-	-	-	-	-	14	87	2,42	1
	>2000 µm	7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	7	87	-	6
M39005-3	250-2000 µm	6	-	-	-	4	-	-	-	-	-	-	-	-	-	-	-	10	41	2,53	1
	>2000 µm	1	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	3	260	-	2
M39006-1	250-2000 µm	3	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	6	42	1,66	1
	>2000 µm	2	-	-	-	1	-	-	3	-	-	-	-	-	-	-	-	6	260	-	6
M39027-1	250-2000 µm	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	-	-	-
M39007-1	>2000 µm	1	-	-	-	-	-	-	-	-	-	2	-	2	-	-	-	7	44	1,59	1
M39018-1	250-2000 µm	-	1	-	-	-	-	9	-	5/3	-	-	-	-	-	-	-	3	116	0,26	1
	>2000 µm	-	2	-	-	-	-	-	-	2/12	-	-	-	-	-	-	-	32	95	5,28	2
M39017-5	250-2000 µm	-	-	-	-	-	-	5	-	-	-	-	-	-	-	-	-	33	173	-	4
	>2000 µm	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	7	81	0,94	1
	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	2	260	-	1
M39028-3	250-2000 µm	-	-	-	-	-	-	1	-	-	-	-	-	-	-	-	-	0	-	-	-
	>2000 µm	-	-	-	-	-	-	-	-	-	-	-	-	3	-	-	-	15	47	3,49	1
M39008-4	250-2000 µm	2	8	-	2	1	-	-	-	-	6	-	-	1	-	-	-	3	93	-	1
	>2000 µm	-	1	-	1/2	-	-	1	-	-	-	-	-	1	-	-	-	40	135	3,86	1
	9	-	1	-	1/6	-	-	2	-	-	-	-	-	1/6	-	-	-	23	260	-	4
M39016-1	250-2000 µm	2	-	-	-	1	-	-	-	-	-	-	-	-	-	-	-	8	-	-	5
	>2000 µm	-	1	-	-	2	-	-	-	-	1	-	-	-	-	1/10	-	11	122	1,36	2
	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	12	260	-	9
M39022-1	250-2000 µm	-	1	-	-	1	-	4	-	-	2	-	2	3	-	-	-	25	68	3,91	1
	>2000 µm	-	1	2	-	-	-	-	-	-	-	-	-	-	-	-	-	6	260	-	2
	6 (1 colonized)	-	-	-	1/14	-	-	-	-	1/20	-	-	-	-	-	-	-	29	-	-	29*
M39009-1	250-2000 µm	1/2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	22	0,52	1
	>2000 µm	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	173	-	1
M39020-1	250-2000 µm	-	-	-	-	2	-	-	-	-	1	-	-	2	-	-	-	9	68	1,51	1
	>2000 µm	-	1	-	-	-	-	-	1	-	-	-	-	2	-	-	-	11	65	-	4
	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	0	-	-	-
M39023-3	250-2000 µm	-	-	-	-	-	-	3	-	-	-	-	-	1	-	-	-	11	36	2,11	1
	>2000 µm	-	1	-	1	-	-	1	1	2/6	-	-	1	4	-	1	-	30	260	-	1
	3	-	-	2	-	-	-	-	-	-	-	-	-	-	-	-	-	4	-	-	3
M39003-1	250-2000 µm	-	-	-	-	-	-	-	-	-	4	-	-	5	2	-	-	15	62	3,29	1
	>2000 µm	-	-	-	2	-	-	3	-	-	8	-	-	2	-	2	-	23	260	-	4
	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	1	-	-	-
M39014-1	1 ship's clinker	-	-	12/20	1/30	-	-	-	-	-	-	-	-	-	-	-	-	47	190	2,48	-
M39013-1	1 outcrop slab	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	22	434	0,51	-
M39010-1	3 outcrop slabs	-	-	-	-	-	-	-	-	1/15	-	-	-	-	-	-	-	88	424	2,07	33*
M39010-2	>2000 µm	-	-	1/5	-	-	-	-	-	-	-	-	-	2/6	-	-	-	24	120	2,00	9
M39021-5	250-2000 µm	-	-	-	-	-	-	1	-	-	-	-	1	2	-	-	-	10	68	2,37	1
	>2000 µm	-	-	4	-	1	-	-	-	-	1	-	2	2	-	-	-	23	260	-	2
	5 (1 colonized)	-	-	12	-	-	-	1	-	-	-	-	-	-	-	-	-	16	-	-	16*
G1K15809	250-1000 µm	-	-	-	-	-	-	-	-	-	1	-	-	-	-	-	-	1	125	0,12	1
	1000-2000 µm	-	-	-	-	-	-	2	-	-	-	-	-	-	-	-	-	2	500	-	1
M39004-1	250-2000 µm	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	12	63	2,64	1
	>2000 µm	-	-	-	1/7	-	-	-	-	-	-	-	-	1	-	-	-	19	260	-	2
M39002-2	250-2000 µm	-	-	-	-	-	-	-	-	-	2	1	3	-	-	-	-	9	62	1,83	1
	>2000 µm	-	-	-	-	-	-	5/2-5	-	-	-	1	1	-	-	-	-	10	260	-	6
	5	-	-	-	1/8	-	-	-	-	-	-	-	-	-	-	-	-	1	-	-	1
M39029-3	250-2000 µm	-	-	5	-	-	-	5	-	-	-	-	-	-	-	-	-	1	52	2,10	1
	>2000 µm	-	-	-	-	-	-	2	-	-	-	-	-	-	-	-	-	5	260	-	2

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